Ljiljana ANDRIJEVIĆ¹ Ilija ANDRIJEVIĆ²



The effect of nucleoside analogues on biochemical parameters in rats' sera

BACKGROUND: Nucleoside analogues are new chemotherapeutic agents which showed activity against various cancer cells in vitro and in vivo. They modulate signal transduction pathways causing growth inhibition, differentiation, apoptosis and modulation of gene expression through distinct mechanisms of action. Tiazofurin is a synthetic analogue of purine nucleosides with modification on C6 atom, while 8-CI-cAMP belongs to C8 analogue group. In our study, we examined the effect of two nucleoside analogues, tiazofurin and 8-CI-cAMP, on biochemical parameters in rats' sera.

METHODS: The experiments were carried out on adult Wistar rats. Tiazofurin and 8-Cl-cAMP were administered IP at a single dose (50 mg/kg/day) and four hours later the rats were sacrificed. In the sera of untreated and treated rats the following parameters were determined: urea, creatinine, uric acid, glucose, cholesterol, triglycerides, albumin, aspartate aminotransferase, alanine aminotransferase, creatine kinase, lactate dehydrogenase, γ -glutamyltransferase, alkaline phosphatase, alfa-amilase, calcium, phosphate and iron.

RESULTS: The results showed that there were no significant statistical differences among biochemical parameters in the sera of treated animals in comparison to untreated rats.

CONCLUSION: These findings implicate that tiazofurin and 8-CI-cAMP did not express toxic effects as measured by parameters above.

¹INSTITUTE OF ONCOLOGY SREMSKA KAMENICA, SREMSKA KAMENICA, YUGOSLAVIA ²INSTITUTE FOR LUNG DISEASES, SREMSKA KAMENICA, YUGOSLAVIA **KEY WORDS:** Antineoplastic Agents; Purine Nucleoside+analogs and derivates; Cyclic AMP; Blood Chemical Analysis; Rats, Wistar

Archive of Oncology 2001,9(3):155-159©2001,Institute of Oncology Sremska Kamenica, Yugoslavia

INTRODUCTION

he disturbance of homeostasis of endogenous nucleosides represents a particular field of scientific interest, because a lot of synthetic analogues become more important potential therapeutics in human oncology (1-7).

The pharmacological approach in the synthesis of novel drugs suggests that the use of purine nucleoside analogues in which heterocyclic structure or sugar moiety is altered in such a way that causes toxic effect when incorporated in different part of the cell. Various compounds used for chemotherapy differ in their chemical structure and mechanism of action (8,9). Widely used nucleoside analogues have substituents on purine or pirimidine ring, which do not exist in natural form. They change the pattern of base pairing or interaction of nucleotide with specific enzymes.

$2-\beta$ -D-ribofuranosyl thiasole-4-carboxamide (tiazofurin)

Many studies have shown that certain imidazol nuclosides like ribavirin, which are potent inhibitors of guanine nucleotide synthesis, have a significant antiviral activity. Assuming that selective regulation of these synthetic pathways in nucleic acids biosynthesis may have an important chemoterapeutic application a series of thiasole-C-nucleosides with ribavirin-like structure, tiazofurin being one of them, were synthesized (10).

Tiazofurin is structural analogue of purine nucleosides with modification on C6 atom. Tiazofurin enters the cell through a simplified nucleoside transport mechanism and joins the cell metabolism. Tiazofurin is a pro-drug that undergoes metabolic conver-

Address correspondence to:

Dr. Ljiljana Andrijević, Institute of Oncology Sremska Kamenica, Institutski put 4, 21204 Sremska Kamenica, Yugoslavia

The manuscript was received: 05. 09. 2001.

Provisionally accepted: 18. 10. 2001.

Accepted for publication: 29. 10. 2001.

^{© 2001,} Institute of Oncology Sremska Kamenica, Yugoslavia

Andrijević Lj.

sion into adenindinucleotide analogue, tiazol-4-karboxamid adenindinucleotide (TAD), active metabolite (11). After that, enzyme adenosine kinase catalyses the conversion of tiazofurin into tiazofurinmonophosphate (TMP) which undergoes conversion into TAD in the presence of NAD pirophosphorilase. TAD is an active metabolite as well as potent inhibitor of enzyme inosine monophosphate dehydrogenase (IMPDH) (12). Inhibition of IMPDH decreases both DNA and RNA synthesis and proliferative ability of malignant cells (Natsumeda et al. 1989). TAD has stronger affinity for NAD/NADH binding site (10-⁷ M) in IMPDH molecule than endogenous coenzyme NAD (13).

There is a direct proportion between the degree of cellular import of tiazofurin, its conversion into TAD in target cells and its pharmacological effect (13).

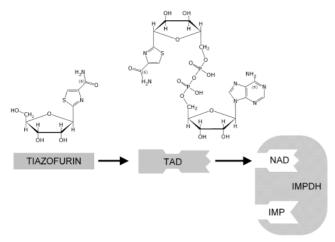


Figure 1. Tiazofurin mechanism of action

Studies of tissue distribution of tiazofurin and its metabolism in a great number of animal models (rodents, rabbits, and dogs) show a significant penetration of tiazofurin in many tissues (14).

In vitro studies of tiazofurin transport through humane erythrocyte membrane show that this molecule is transported by endogenous nucleosides transport system (15).

8-Chloro-cyclic adenosine 3'5' monophosphate (8-Cl-cAMP)

8-Chloro-cyclic adenosine 3'5' monophosphate belongs to C8 analogue group; with modification on C8 atom of purine ring (Figure 2).

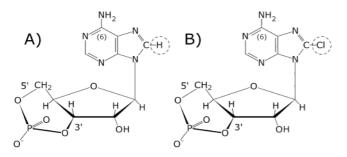


Figure 2. The structure of cyclic 3 '5' - AMP (A); and 8-CI-cAMP (B)

None of so far examined C2, C6 or C8 analogue of cAMP, showed such a great regulatory effects on the growth inhibition of human cancerous cell lines spectrum, as 8-CI-cAMP did (16).

The mechanisms of normal differentiation in cancer cells are disturbed, but could be restored by action of 3⁵ monophosphate analogues through specific receptors (17).

Cyclic AMP, as the intracellular regulating factor, plays a key role in differentiation and growth regulation in different cell types; however, the exact mechanism of action is still unknown (18). In mammal cells cAMP is acting via cAMP dependent protein kinase (PKA) receptors PKA I and PKA II (19,20).

The mechanism of action of 8-CI-cAMP is still unknown. It is supposed that inhibitory growth effect of 8-CI-cAMP might be related to decreased ratio of PKA I/PKA II isoforms in cancer cells (21). However, some results suggest that inhibitory growth effect of 8-CI-cAMP could be mediated by its metabolite 8-CI-adenosine as well (22).

The aim of this study was to examine the effect of two nucleoside analogues, tiazofurin and 8-CI-cAMP, on biochemical parameters in rats' sera.

MATERIALS AND METHODS ____

Animals. Thirty Wistar rats, male and female, 250-300 g weight, bred at the Farm for Experimental Animals, ICN Yugoslavia Institute, were used in this study.

Agents. Tiazofurin and 8-CI-cAMP were obtained from the ICN Yugoslavia Institute.

Application of tiazofurin and 8-CI-cAMP. Tiazofurin and 8-CI-cAMP were administered to Wistar rats intraperitoneally (IP) in a single dose (50 mg/kg/day). Four hours after administration of the agents, blood samples were taken by cardiac puncture. After that rats were sacrificed.

Determination of biochemical parameters. In the sera of untreated and treated rats, the following parameters were determined: urea, creatinine, uric acid, glucose, cholesterol, triglycerides, albumin, aspartate aminotransferase, alanine aminotranferase, creatine kinase, lactate dehydrogenase, γ -glutamyltransferase, alkaline phosphatase, alpha-amylase, calcium, phosphate and iron (Table 1). We used EXPRESS 550 CIBA CORNING analyzer to determine the concentrations of biochemical parameters in rats' sera.

Statistics. The results are expressed as means (standard deviation (SD). Student's *t* test was used for the statistical analysis.

^{© 2001,} Institute of Oncology Sremska Kamenica, Yugoslavia

Table 1. The biochemical parameters, their units and methods for the quantitative determination in rats' sera

Biochemical parameters	Methods
1. Urea (mmol/L)	Urease/glutamate-dehydrogenase
Creatinine (µmol/L)	Alkaline-picrate (Jaffe)
3. Uric Acid (µmol/L)	Uricase- peroxidase
4. Glucose(mmol/L)	Glucose oxidase
5. Cholesterol (mmol/L)	Cholesterol oxidase
Triglycerides(mmol/L)	Lipase/glycerol
7. Albumin (g/L)	Bromcresol green
8. Aspartate aminotransferase (U/L)	AST, pyridoxal 5- phosphate
Alanine aminotransferase (U/L)	ALT, pyridoxal 5- phosphate
10. Creatine kinase (U/L)	CK – NAC (NAC=activator: N-acetylcysteine)
11. Lactate dehydrogenase (U/L)	Pyruvate, > 0.7 mmol/L
12. γ-Glutamyltransferase (U/L)	γ-glutamyl-3carboxy-4-nitroanilide
13. Alkaline phosphatase(U/L)	p-nitrophenyl phosphate
14. α-Amylase (U/L)	Maltoheptaoside
15. Calcium (mmolL)	Orto-cresolphtaleine
16. Phosphate(mmol/L)	Phosphomolybdate
17. Iron (µmol/L)	Colorimetric test without deproteinisation

RESULTS

Because biochemical parameters have not been evaluated so far in sera of Wistar rats, we determined their concentrations in control samples. The control group consisted of ten rats' sera samples (five males and five females). In the sera of untreated and treated rats the following parameters were determined: urea, creatinine, uric acid, glucose, cholesterol, triglycerides, albumin, aspartate aminotransferase, alanine aminotranferase, creatine kinase, lactate dehydrogenase, γ -glutamyltransferase, alkaline phosphatase, alfa-amilase, calcium, phosphate and iron (Table 2).

Table 2. The concentrations of biochemical parameters in control group expressed as a range of two standard deviations

Biochemical parameters	Control group	
	male	female
1. Urea (mmol/L)	4.4-8.4	6.6-7.2
2. Creatinine (µmol/L)	57-92	78-82
3. Uruc Acid (µmol/L)	32.7-71.3	19.7 – 138.3
4. Glucose (mmol/L)	7.1-7.8	4.6-8.8
5. Cholesterol (mmol/L)	2.14-3.1	2.45-3.11
6. Triglycerides (mmol/L)	1.67-2.19	0.68-2.6
7. Albumin (g/L)	36-62	31-59
Aspartate aminotransferase (U/L)	129-142	93-227
Alanine aminotransferase (U/L)	50-66	35.8-83.4
10. Creatine kinase (U/L)	522-938	599-997
 Lactate dehydrogenase (U/L) 	1366-2036	597-3023
 γ-Glutamyltransferase (U/L) 	23.4-38.2	26.6-31.8
Alkaline phosphatase (U/L)	461-657	279-593
14. α-Amylase (U/L)	3611-5984	2721-5607
15. Calcium (mmol/L)	1.95-3.05	2.15-2.85
16. Phosphate (mmol/L)	1.71-2.17	1.56-2.08
17. Iron (μmol/L)	32.2-41.92	33.12-59.76

The group treated with tiazofurin consisted of ten rats (five males and five females). Four hours after administration of a single dose of tiazofurin we determined the concentrations of biochemical parameters in rats' sera (Table 3).

The group treated with 8-CI-cAMP consisted of ten rats (five males and five females). Four hours after administration of a single dose of 8-CI-cAMP we determined concentration of biochemical parameters in rats' sera (Table 4).

Table 3. The mean values of biochemical parameters in rats' sera after administration of single dose of tiazofurin

Biochemical parameters	Treated rats Tiazofurin	
	male	female
1. Urea (mmol/L)	6.1	5.82
2. Creatinine (µmol/L)	66.16	69.18
3. Uruc Acid (µmol/L)	41	61
4. Glucose(mmol/L)	7.5	6.5
5. Cholesterol (mmol/L)	2.18	2.76
6. Triglycerides(mmol/L)	1.65	1.14
7. Albumin (g/L)	39	42
Aspartate aminotransferase(U/L)	145	138
Alanine aminotransferase(U/L)	75.3	52.8
10. Creatine kinase (U/L)	730	750
 Lactate dehydrogenase (U/L) 	1664	1780
12. γ-Glutamyltransferase (U/L)	24.4	27.6
 Alkaline phosphatase(U/L) 	551.4	411.8
14. α-Amylase (U/L)	5792.2	4802.6
15. Calcium (mmol/L)	2.5	2.46
16. Phosphate(mmol/L)	1.92	1.7
17. Iron (µmol/L)	39.18	49.46

Table 4. The mean values of biochemical parameters in rats' sera after administration of single dose of 8-CI-cAMP

Biochemical parameters	Treated rats 8-CI-cAMP	
	male	female
1. Urea (mmol/L)	7.4	6.78
2. Creatinine (µmol/L)	74.12	76.2
3. Uruc Acid (µmol/L)	71	84
4. Glucose(mmol/L)	6.1	6.3
5. Cholesterol (mmol/L)	2.34	2.8
6. Triglycerides(mmol/L)	1.42	1.15
7. Albumin (g/L)	41	42
Aspartate aminotransferase (U/L)	204	174
Alanine aminotransferase (U/L)	74.5	61.5
10. Creatine kinase (U/L)	770	831
 Lactate dehydrogenase(U/L) 	2267	2354
 γ-Glutamyltransferase (U/L) 	43.4	35.2
Alkaline phosphatase(U/L)	574	502.4
14. α-Amylase (U/L)	6140	4385.8
15. Calcium (mmol/L)	2.57	2.44
16. Phosphate(mmol/L)	2.02	1.74
17. Iron (μmol/L)	40.82	46.6

DISCUSSION ____

We examined the effect of two nucleoside analogues, tiazofurin and 8-CI-cAMP, on biochemical parameters in rats' sera. To our knowledge, the biochemical parameters (enzymes, metabolites and oligoelements) have not been determined in Wistar rats' sera so far. Our aim was to determine their concentrations prior to treatment. In the sera of ten untreated rats the following parameters were determined: urea, creatinine, uric acid, glucose, cholesterol, triglycerides, albumin, aspartate aminotransferase, alanine aminotranferase, creatine kinase, lactate dehydrogenase, y-glutamyltransferase, alkaline phosphatase, alfa-amilase, calcium, phosphate and iron (Table 2). After determination of control values of biochemical parameters, we determined the concentrations of the same parameters after administration of a single dose of tiazofurin and 8-CI-cAMP. Both drugs were applied at doses that induce antitumor and/or antiproliferative effects on C6 cell line (24-26). The mean values of all investigated biochemical parameters after administration of tiazofurin were within range of

Andrijević Lj.

two standard deviations compared to control values, except for creatinine. The difference between concentration of creatinine in treated female and creatinine concentration in untreated female rats was statistically significant (p < 0.05; Table 3). The inhibition of IMPDH leads to the decrease of guanilate synthesis, depletion of guanilate in cells and reduction of DNA and RNA synthesis, which causes the reduction of malignant cells' proliferative ability (5). Antitumor effect of tiazofurin is shown in vitro and in vivo in a number of solid tumors. Except for anti-proliferative effect, tiazofurin also leads to differentiation of breast cancer cells - MCF-7, ovarian carcinoma cells OVAR-5, and differentiation and apoptosis of leukemia cell line - K-562 (23). Examination of antitumor effect of tiazofurin in human glioma line has shown that tiazofurin has significant antiproliferative and differentiation potential, but it has no cytotoxic effect on these cells (24). The mean values of all investigated biochemical parameters after administration of 8-ClcAMP were within the range of two standard deviations as compared to control values, except for glucose. The difference between concentration of glucose of treated males and untreated males was statistically significant (p < 0.05; Table 4). 8-CI-cAMP showed an inhibitory effect to breast cancer and colon cancer cell line growth, without toxic signs (27). It is showed that 8-CI-cAMP significantly inhibits proliferation of stomach cancer cells (19). Application of previously examined cAMP analogues, such as dibutiril-cAMP or medicines that increase cAMP levels in cells, led to controversial opinions about the cAMP effect on cancer cells growth. 8-CI-cAMP effect to inhibition of growth was compared with dibutiril-cAMP effect on wide spectrum of human cancer cell lines (21).

The anti-cancer compound tiazofurin is currently being tested in patients (Phase II/III clinical trials) with chronic myelogenous leukemia (CML) in accelerated phase or blast crisis, ovarian cancer, and multiple myeloma. (28).

Experimental results obtained *in vivo* and *in vitro* with 8-CI-cAMP make basis for clinical investigation of the compound in the therapy of human glioma (26).

CONCLUSION

Our results showed that there were no significant differences among investigated biochemical parameters in the sera of treated animals by tiazofurin and 8-CI-cAMP in comparison to untreated rats. These findings implicate that tiazofurin and 8-CI-cAMP do not express toxic effect. This is an important conclusion in our further investigation of tiazofurin and 8-CI-cAMP in clinical treatment.

Acknowledgments

We would like to thank Assist. Prof. Aleksandar Kerenji for critical reading of the manuscript and Dimitar Jakimov for excellent technical asisstance.

REFERENCES

- Puiroud S, Pinard E, Seylaz J. Dynamic cerebral and systemic circulatory effects of adenosine, theophylline and dipyridamole. Brain Res 1988;453:287-98.
- Daval J L, Nehlig, Nicolas F. Physiological and pharmacological properties of adenosine: therapeutic inplications. Life Science 1991;49:1435-53.
- Henderson J F, Smith C M, Zombor G. Regulation of purine metabolism in lymphocytes. Procceding of the society for experimental biology and medicine 1985;179:419-26.
- Weber G, Natsumeda Y, Lui MS. Control of enzymic programs and nucleotide pattern in cancer cells by acivicin and tiazofurin. Adv Enzyme Regul 1984;22:69-73.
- Natsumeda Y, Ikegami T, Olah E, Weber G. Significance of purine salvage in circumventing the action of antimetabolites in rat hepatoma cells. Cancer Res 1989;49:88-92.
- Plagemann PGW, Wohlhueter RM, Woffendin C. Nucleoside and nucleobase transport in animal cells. Biochemica and Biophysica Acta 1988;947:405-43.
- Jackson RC, Harkrader RJ. The contributions of de novo and salvage pathways of nucleotide biosynthesis in normal and malignant cells. In: Tattersall MHN, Fox RM, eds. Nucleosides and cancer treatment. Sydney: Academic Press; 1981. p. 18-31.
- Flowers A, Levin V. Chemotherapy of brain tumors. In: Kye HA, Laws ER, Jr, eds. Brain tumors. Edinburg: Churchile livingstone; 1995. p. 249-60.
- Tricot G, Weber G. Biochemically targeted therapy of refractory leukemia and mieloyd blast crisis of chronic granulocytic leukemia with tiazofurin, a selective blocker of inosine 5-phosphate dehydrogenase activity. Anticancer Res 1996;16:3341-8.
- Srivastava PC, Pickering MV, Allen LB, Streeter DG, Campbell MT, Witkowski JG et al. Synthesis and antiviral activity of certain thiasole c-nucleosides. Med Chem 1977;20:256-62.
- 11. Crabtree GW. Tiazofurin summary of information. Viratec Inc. Costa Mesa, Ca, 1988.
- **12.** Weber G, Praida N. Targeted and non-targeted actions of anti-cancers drugs. Adv Enzime Regul 1994;34:71-89.
- Yamada Y, Natsumeda Y, Webwr G. Action of the active metabolites of tiazofurin and ribavirin on purified IMP dehydrogenase. Biochemistry 1988;27:2193-6.
- Arnold TC, Jayaram NH, Harper RG. Disposition and metabolism of tiazofurin in rodents, rabbits and dogs. Drug Metabolism and Disposition 1984;12:165-73.
- Monks A, Marqez V, Mao DT, Cysyc RL. Uptake of2-(-D-ribofuranosyl thiasole-4-carboxamide (tiazofurin) and analogues by the facilitated transport mechanism of erythrocytes. Cancer Letters 1985;28:1-8.
- Cho-Chung YS, Clair T, Tagliaferri P, Ally S, Katsaros D, Tortora G, et al. Siteselective cyclic AMP analogs as new biological tools in growth control, diferentation and proto-oncogene regulation. Cancer Invest 1989;7:161-77.
- Cho-Chung YS. Role of cAMP receptor proteins growth, diferentation and supression of malignancy: new approaches to therapy. Cancer Res 1990;50:7093-100.

- Ally S. Inhibition of growth and modulation of gene expression in human lung carcinoma in athymic mice by site-selective 8-CI-cyclic adenosine monophosphate. Cancer Res 1989; 49: 5650-5.
- **19.** Katsaros D. Site-selective cyclic AMP analogs provide a new approach in the control of cancer cell growth.Febs Lett.1987; 223: 97-103.
- Adams SR, Harotunian AT, Buechler YJ. Fluorescence ratio imaging of cyclic AMP in single cells. Nature 1991; 349:694-7
- **21.** Tortora G, Ciardillo F, Ally S. Site selective 8-chloro- adenosine 3,5 monophosphate inhibits transformation and transforming grwoth factor alpha production in Ki-ras transformed fibroblasts. FEBS 1989; 242:363-7.
- 22. Van Lookeren Campagne MM, Diaz FV, Jastorff B, Kessin RH. 8-chloro adenosine 3,5-monophosphate inhibits the growth of Chinese hamster ovary and molt-4 cells trough its adenosine metabolite. Cancer Res 1991;51:1600-5.
- **23.** Olah E, Csokay B, Praida N. Molecular mechanism in the antiproliferative action of taxol and tiazofurin. Anticancer Res 1996; 16:2469-78.
- 24. Drabek K. Antitumorous effects of tiazofurin to humane glioma cell lines . M.Sc. thesis. Belgrade: Medical Faculty; 1998.
- **25.** Jovanovic S. Transport of endogenous nucleosides and their synthetic analogs throuth the blood tumor barier in rats.Ph.D. thesis. Belgrade:Medical Faculty; 1996.
- **26.** Andrijevic Lj. The influence of tiazofurin and 8-CI-cAMP on tumor marker on experimental glioma rats. Ph.D. thesis. Belgrade:Medical Faculty; 1999.
- Bianco C, Tortora G, Baldassare G. 8-Chloro-cyclic AMP inhibits autocrine and angiogenic growth factor production in human colorectal and breast cancer. Clinical Cancer Res 1997; 3:439-48.
- 28. Available from: URL http:// www.icnpharm.com