



Nevenka STANOJEVIĆ-BAKIĆ
Ljiljana VUČKOVIĆ-DEKIĆ

Interferons in the therapy of solid tumors.

Part III. Interferon and various solid malignancies

ABSTRACT

Interferons exert a consistent therapeutic effect in a proportion of patients with renal cell carcinoma and melanoma. In other solid malignancies, this therapeutic approach is investigated at more limited extent; therefore, it is still in experimental area. In this review, we analyzed the clinical trials that used the IFN as monotherapy or, more frequently, as combined biochemotherapeutic regimen. This therapeutic strategy was not justified in colon cancer. Similarly, IFNs did not make a major progress in the treatment of lung cancer regardless the tumor type. Very limited activity was seen also in advanced breast cancer. In most other solid tumors, clinical experience is insufficient and only anecdotal benefits were reported. This is equally true for premalignant lesions, the possibilities of IFN-therapy of which are largely overlooked. In the future, some approaches such as combination of IFN and hormonotherapy in breast cancer, and with retinoic acid in squamous cell carcinomas, deserve further investigation. The optimization of IFN regimens in solid malignancies is the aim of current efforts. Better understanding of biological mechanisms of specific tumor sensitivity, and also the mechanisms of resistance of sensitive tumor types to IFN, will probably lead to the defining features of tumor responsiveness.

Key words: Colon cancer; Lung cancer; Breast cancer; Immunotherapy

INSTITUTE FOR ONCOLOGY AND RADIOLOGY OF SERBIA, BELGRADE, YUGOSLAVIA

Archive of Oncology 2000,8(2):65-72©2000, Institute of oncology Sremska Kamenica, Novi Sad, Yugoslavia

INTRODUCTION

Although at the first glance the results of interferon (IFN) therapy of solid tumors might seem discouraging, some metastatic tumors such as renal cell carcinoma (RCC) (1) and malignant melanoma (MM) (2) undergo regression in a fraction of treated patients. Apart from these tumors, IFNs were tried in a wide range of other solid malignancies. In the therapy of common solid tumors such as colon and lung cancers, the use of IFN is still experimental. In this review we summarize the results of clinical

studies that used IFN, alone or in combination with other oncological treatment modalities, in the therapy of various cancers.

Colorectal cancer

There is, presently, no satisfactory standard treatment for advanced colorectal carcinoma (CC). Most commonly used chemotherapeutic agent, 5-fluorouracil (5-FU), definitely has some, but only modest activity, giving the response rates of 10-15% and no consistent effect on survival. Because of that, the developments of new therapeutic strategies are ongoing, with the aim to improve the response rate, time to progression and survival. Many of them include IFN, which was stimulated by preclinical data and encouraging results obtained in RCC and MM patients.

IFN as a single agent therapy

Several clinical trials used IFN α monotherapy in advanced colorectal cancer; a response rate of only 2% was obtained, regardless dose or schedule (3). Therefore, such a therapy seems to have no activity and virtually no clinical effects.

Few studies with IFN β (4) or IFN γ (5) also showed very limited activity. These disappointing results were the reason for abandonment of the IFN single-agent therapy of advanced CC.

The very poor responsiveness of this gastrointestinal malignancy to IFN treatment may be ascribed, at least in part, to the presence of

both specific and non-specific IFN inactivators/inhibitors in sera of the patients (6,7); the nature of these factors is still unclear.

IFN and chemotherapy drug combinations

IFN and 5-FU combination. Preclinical data suggesting that IFN α and 5-FU have a synergistic cytostatic effect upon cultured colon carcinoma cells (8), stimulated an increasing number of clinical investigations to evaluate the therapeutic potential of this combination. This was also supported by the observation that IFN-chemotherapy combination may reduce the IFN-inhibitors/inactivators in cancer patients (9). The results of 25 selected trials are presented in Table 1 (10-32). In various treatment schedules, 5-FU was used at doses ranging from 225-750mg/m² (usually 750mg/m²), and IFN α at doses of 0,5 - 20MU/day. The overall response (OR) rates varied very much (3-70%), giving an average OR of 27%. These divergent therapeutic results may be accounted, at least in part, to the small number of patients in some trials, and inclusion of patients refractory to 5-FU monotherapy. It is evident that the initial impressive response rate (63%) obtained in the Wadler's report (11) has failed to be reproduced; the only exception is the recent study of Kimm et al. (28), the relevance of which is hampered by the small number of patients. However, the overall response rate of about 30%, although modest, is still higher than that of the each agent

Abbreviations used in text: IFN - interferon; RCC - renal cell carcinoma; MM - malignant melanoma; CC - colorectal cancer; 5-FU - 5-fluorouracil; LV - leucovorin; CR - complete response; PR - partial response; OR - overall response; NSCLC - non-small cell lung cancer; SCLC - small cell lung cancer; SCC - squamous cell carcinoma; IL-2 - interleukin 2; sIL-2R - soluble IL-2 receptors; ChT - chemotherapy; RT - radiotherapy; RA - retinoic acid; BRMs - biological response modifiers

Address correspondence to:

Dr. Nevenka Stanojević-Bakić, Institute for oncology and radiology of Serbia, Pasterova 14, POB 228, 11000 Belgrade, Yugoslavia

The manuscript was received: 22. 02. 2000.

Provisionally accepted: 28. 02. 2000.

Accepted for publication: 10. 03. 2000.



monotherapy. In terms of survival, the results of several randomized trials gave no indication that the IFN α -5-FU combination had any advantage over the 5-FU monotherapy.

It should be noted that the constant evidence from most clinical trials was the increased systemic toxicity of 5-FU by IFN α , requiring the reduction of IFN dose in some patients (33).

Therefore, although these two drugs' synergism was reported in several preclinical studies, the results of trials analyzed in this review did not justify this therapeutical strategy. The enhanced toxicity, no survival benefit, along with cost consideration, compromise the beneficial effect on the OR rate.

Similarly to IFN α , IFN β also potentiates the antitumor activity of 5-FU against human colon cancer cells *in vitro* and *in vivo* (34). Based on these findings, three recent clinical studies used IFN β for the 5-FU biomodulation (Table 1).

The mean OR rate of 27% is similar to that of

IFN α +5-FU combination. It seems that IFN β does not increase the toxicity of 5-FU (30,32), and that this combination is less toxic than that of IFN α +5-FU. A significant increase of survival was reported in one study (32), suggesting that this combination might deserve further investigation.

IFN+5-FU+LV combination

Another strategy aiming to enhance cytotoxicity of 5-FU is focused on the use of triple drug combination - 5-FU, leucovorin (LV) and IFN α . The potential advantage might be double modulation of 5-FU by LV and IFN, the drugs with different mechanisms of action. The results of 15 clinical trials with 908 patients are presented in Table 2 (35-43). The average OR of 29% was obtained with different doses and schedules of these drugs' administration (range 10-54%). The response rate and survival duration were not significantly better than that obtained

Table 2. Response of advanced colorectal cancer patients to the IFN+5-FU+LV combinations.

Study	Evaluable patients	CR	PR	OR (%)	Mean survival (months)
Recchia 1992*	32	2	5	22	
Schmoll 1992*	43			10	
Cascini 1992*	45			51	
Sobrero 1992 ³⁵	15			20	
Pensel 1993*	24	4	6	41	
Kocha 1993*	240	7	43	21	
Grem 1993 ³⁶	44	3	21	54	
Pun 1993 ³⁷	45			25	11
Seymour 1994*	83			30	
Pazdur 1994 ³⁸	47	3	11	30	
Kosmidis 1996 ³⁹	51			10	7.2
Tournigand 1997 ⁴⁰	50	1	21	44	25
Kohne 1997 ⁴¹	33			15	9.9
Kohne 1998 ⁴²	49			27	19.6
Hausmaninger 1999 ⁴³	107	5	33	36	
Total: 15	908			29	(mean)

* References cited in Raderer³³ and Kjaer³

with 5-FU alone or combined with either LV or IFN. In addition, virtually all trials have demonstrated a significantly higher rate of adverse events, and decreased quality of life. This is because such regimens cannot presently be recommended for routine use outside clinical investigation (43).

There exist little data for IFN and antineoplastic drugs other than 5-FU and LV for patients with advanced colorectal cancer (44,45); they provided no evidence of any therapeutic advantage over the 5-FU regimens.

IFN α -5-FU combination and other biotherapeutics

Further attempts to improve response rate and survival of advanced CC patients introduced IFN γ or interleukin-2 (IL-2) to the combination of 5-FU and IFN α (with or without LV). The rationale for such combinations was the preclinical data suggesting synergy between these biological agents, which was confirmed in clinical studies in RCC and MM patients (1,2). Few trials (46-49) using different scheduling and doses of these biochemotherapeutics, showed that these multiagent combinations failed to improve clinical benefit of 5-FU monotherapy, and were accompanied by severe toxicity that required the treatment interruptions or dose reductions.

As in RCC and MM (1,2), it is not possible to define the responders to IFN combination therapies. The predictors of response are still lacking. It is unlikely that ras mutations (such as c-Ki-ras2, which occurs early in the pathogenesis of CC and is found in about 40% of patients) will have significant prognostic value for either response to therapy or survival (50). Similarly, the immunostimulation obtained by IFN treatment did not predict improved clinical outcome (51).

In conclusion, after the initial hopes, the combination of IFN- α with 5-FU and other biochemotherapeutics does not seem to fulfill the original expectations. It is not sufficiently effective, it is toxic, and it is costly. Further research efforts are required and new treatment strategies are needed if progress is to be

Table 1. Response of advanced colorectal cancer patients to the IFN+5-FU combinations.

Study	Evaluable patients	CR	PR	OR (%)	Mean survival (months)
IFN α+5-FU					
Wrightley 1984*	14	0	2	14	
Clark 1987 ¹⁰	29	0	1	3	
Wadler 1990 ¹¹	32	0	20	63	
Pazdur 1990 ¹²	45	1	15	36	
Kemcny 1990 ¹³	35	0	9	26	
Fornasiero 1990 ¹⁴	21	4	5	43	
Huberman 1991*	33	0	13	39	
Wadler ECOG 1991 ¹⁵	36	1	14	42	
Meadows 1991 ¹⁶	17	2	2	23	
Weh 1992 ¹⁷	55	0	17	31	
Rubio 1992 ¹⁸	33	3	5	24	
Pazdur 1993 ¹⁹	39	1	11	31	
John 1993 ²⁰	18	1	5	33	
York 1993*	121			31	
Findlay 1994 ²¹	118	0	5	8	
Hill 1994*	63			31	11
CORFU-A 1995 ²²	243			21	11
DiConzanzo 1995*	92			9	
Hill 1995 ²³	52			19	8
Kohne 1995*	68			19	
Piga 1996 ²⁴	64	2	4	9	12
Dufour 1996 ²⁵	56	3	8	20	12,3
Patt 1997 ²⁶	45	3	12	33	
Perez 1998 ²⁷	33	0	2	6	5
Kim 1998 ²⁸	10	1	6	70	
Kohne 1998 ²⁹	90			18	12,7
Total: 26	1462			27	(mean)
IFN β+5-FU					
Joffe 1997 ³⁰	21	1	3	19	8,4
Wadler 1998 ³¹	59			28	
Villar-Grimalt 1999 ³²	25			33	15,9
Total: 3	105			27	

* References cited in Kjaer³

obtained. Some attempts in this direction are already ongoing; for instance, an important approach to enhance the efficacy of monoclonal antibodies-based protocols in the therapy of this malignancy (52) utilizes the ability of IFNs to up-regulate carcinoembryonic and other tumor-associated antigens (53).

Lung cancer

Although chemo- and radiotherapy do have activity in lung cancer, the results of these therapeutic options are unsatisfactory. Therapeutic regimens that are currently used in advanced non-small cell lung cancer (NSCLC) yield the average OR rates of 20-30%, while median survival may be as low as 6-8 months. Small-cell lung cancer (SCLC), which distinguishes itself from NSCLC by more aggressive clinical course and median survival less than 3 months in the absence of treatment, has greater responsiveness to chemotherapy, but median survival remains 12-14 months.

In the search of new systemic strategies against lung cancer, biologic agents such as IFNs have been reconsidered for the treatment programs. Clinical studies that include IFNs are heterogeneous in regard to the IFN type and other biochemotherapeutics used in combination therapy; they frequently dealt with small sample series, and patients groups are often unmatched by stage of disease and category of responses.

IFNs in advanced non-small cell lung cancer (NSCLC)

The IFN monotherapy is inactive in this disease; the responses were rare and no impact on survival was seen (54).

Clinical experiences with IFNs as adjunctive treatment of NSCLC are limited. The summarized data of trials that used IFN in combination with other agents are presented in Table 3 (55-72). It is evident that addition of IFNs to conventional chemotherapy confers little or no benefit; these combinations are usually accompanied by increased toxicity. When IFNs were combined with radiotherapy (RT), different results have been reported. The IFN β -therapy preceding RT gave encouraging response rate in the McDonald's study (65); concurrent treatment with IFN α and RT did not provide any advantage over RT alone (64), while the concomitant treatment of NSCLC patients with IFN γ and fractionated thoracic radiation was associated with severe, life-threatening toxicity without effect on survival (66).

In the studies using IFN in combination with retinoic acid (RA) or IL-2, no antitumor response was seen (Table 3). Multiagent regimens including chemotherapy, IFN and the thymic preparation thymosin- α 1, gave an improved response rate (71,72), which was associated with reduced toxicity. These promising results need to be confirmed in larger randomized trials.

Table 3. Response of advanced non-small lung cancer patients to IFN in combination with standard oncologic therapy and various biotherapeutics.

Study	Therapy	Evaluable patients	CR	PR	OR (%)	Median survival (months)
Schiller 1989 ⁵⁵	IFN γ +IFN β + ChT	18	0	2	11	8
Bowman 1990 ⁵⁶	IFN α +ChT	60	0	18	30	
Rosell 1991 ⁵⁷	IFN α +ChT	30			13	
Lind 1991 ⁵⁸	IFN α +ChT	45	2	7	20	
Ardizzoni 1993 ⁵⁹	IFN α +ChT	90			19	5,5
Halme 1994 ⁶⁰	IFN γ +ChT	27	0	8	29	6-7
Quan 1994 ⁶¹	IFN γ +IFN α +ChT	27	0	9	35	6-7
Quan 1996 ⁶²	IFN α +ChT	6	0	2	33	
Prior 1999 ⁶³	IFN γ +ChT	18	0	7	39	
Maasilta 1992 ⁶⁴	IFN α +RT	32	0	5	15	11
McDonald 1993 ⁶⁵	IFN β +RT	10	0	6	69	
Show 1995 ⁶⁶	IFN γ +RT	32	14	12	81	7-8
Krigel 1991 ⁶⁷	IFN β +IL-2	18	1	2	4	9
Rinaldi 1993 ⁶⁸	IFN α +RA	37	0	1	4	
Arnold 1994 ⁶⁹	IFN α +RA	34	0	1	3	
Athanasiadis 1995 ⁷⁰	IFN α +RA	25	2	2	16	14
Garaci 1995 ⁷¹	IFN α +ChT+T α 1	56	2	22	43	12,6
Salvati 1996 ⁷⁷	IFN α +ChT+T α 1	11			33	

ChT-chemotherapy; RA-retinoic acid; T α 1-thymosin α 1; RT-radiotherapy

IFNs in small-cell lung cancer (SCLC)

Similarly to NSCLC, the IFN trials in SCLC are heterogeneous, which makes the interpretation of the results difficult. The available data on this matter are presented in Table 4 (73-83). In most studies, IFN α was used as maintenance therapy for patients in whom complete (CR) or partial responses (PR) were achieved by induction chemo- and radiotherapy. In contrast to Mattson et al. (74,75), whose results were significantly in favor of IFN α therapy (in patients for whom other prognostic factors were favorable), other reports were negative or inconclusive (76,81,82). In the trials that used IFN γ , no beneficial effect on survival was observed (78-80). It is noteworthy that IFN-therapy was usually associated with toxic effects, which frequently required the discontinuation of treatment.

Presently, it seems that IFNs do not make a major progress in the treatment of lung cancer. Further investigations to define the active biochemotherapeutic combination and optimal dosing schedules are necessary.

Breast cancer

Numerous attempts to improve the efficacy of chemotherapy of metastatic breast cancer by

the additions to, and substitutions of one or other chemotherapeutic agent/s in conventional regimens, have failed to produce further substantial improvement of response rate or response duration. Because of that, the biotherapy, including IFNs, in a combination strategy against breast cancer has been tried. However, limited number of clinical trials are available thus far and this approach is still in experimental area.

Encouraging data of initial reports concerning therapeutic potential of IFN α , have not been confirmed in subsequent clinical trials (84-88): in most patients, IFN therapy had negligible activity.

In few clinical studies, IFN α was used in combination with IL-2; minor objective response associated with considerable toxicity was reported (89-91).

In patients with subcutaneous metastases, the intralesional therapy with IFN α +IFN γ has been tested in two small trials. Promising locoregional antitumor activity associated with extensive immunomodulation was found (92,93), but further follow-up studies are needed to confirm these results.

Another combination of biotherapeutics - IFN α and thymostimulin, was evaluated in



Table 4. Interferons in small-cell lung cancer patients responsive to induction chemotherapy.

Study	IFN type	Number of patients	Median time to progression (months)	Median survival (months)	2-year survival (%)
Kohne 1992 ⁷³	IFN- α	25	13.5	16.5	
	control				
Mattson 1992 ⁷⁴	IFN- α	91		11	18*
	ChT	59		11	7
	control	87		10	6
Mattson 1997 ⁷⁵	IFN α	91			10«
	ChT	59			2«
	control	87			2«
Kelly 1993 ⁷⁶	IFN- α	64	9	13	35
	control	68	10	16	35
Glisson 1993 ⁷⁷	IFN- α	14		10	
Jett 1994 ⁷⁸	IFN- γ	51	6,9	13,3	27 ^{ns}
	control	49	8,1	18,8	33
Bitran 1995 ⁷⁹	IFN- γ	41	3,6		
van Zandwijk 1997 ⁸⁰	IFN- γ	59		8,9	17
Tummarello 1997 ⁸¹	IFN- α	14	12	15	28 ^{ns}
	control	12	7	9	25
# Zarogoulidis 1996 ⁸²	IFN- α +ChT	42		11,3	
	ChT	39		10	
# Prior 1997 ⁸³	IFN α +ChT	43	7,6		14
	ChT	34	5,4	**	0

* $p < 0,01$ in comparison to control** $p < 0,02$ in comparison to control

ns - not significant in comparison to control

IFN concomitant with chemotherapy

ChT - chemotherapy

« 5 year survival

advanced breast cancer by Munno et al. (94). In clinical terms, patients administering this combination could complete chemotherapeutic cycles without interruptions; they had fewer infections in comparison to patients receiving different therapeutic regimen.

Generally, all these data show that IFNs, used either as single agent or in combination with other biotherapeutics, have no or very limited clinical activity in advanced breast cancer.

During the last few years, there is an increasing number of trials using IFNs in combination with hormone therapy in the treatment of this malignancy. Such an approach is based on *in vitro* and *in vivo* evidence that IFNs can induce estrogen receptors and reconstitute the sensitivity of mammary carcinoma to tamoxifen (95-97). In the first clinical trial that included 43 tamoxifen-resistant patients, 26% tamoxifen-responders were seen after IFN β pretreatment (98). Similarly, in a small pilot study of Seymour (99), four out of seven patients responded to treatment with IFN α and tamoxifen. However, subsequent clinical studies reported either no substantial improvement of the efficacy of tamoxifen after IFN treatment, or the higher

response in selected patients (predominant soft tissue disease) only (100-102). In a recent randomized study of Barak et al. (103), IFN β and IFN γ combined with hormone therapy were used. Clinical response was correlated with various cytokine levels. A favorable response to the therapy was associated with significant increase of the IFN γ levels. Baseline levels of IFN γ and sIL-2R were found to be prognostic for clinical response, and to be the most sensitive cytokine parameter for defining the clinical utility of the combination of IFNs and hormone therapy in this malignancy.

Based on the synergistic antiproliferative effect of IFN, retinoids and tamoxifen on the breast cancer cell lines (96,104,105), this combination was tested in the studies of Recchia et al. (106-108); such regimen was effective, with acceptable toxicity, as salvage therapy in pretreated advanced breast cancer patients.

Taking altogether, the true value of these combination regimens cannot be recognized until additional information, obtained in larger number of patients, became available. At present, the use of IFNs-tamoxifen combinations in

the treatment of advanced breast cancer remains investigational, and the optimal scheduling is still undetermined.

Either alone or combined with different biotherapeutics, IFN has also been tried, although less frequently, in tumors other than the above-mentioned ones. In several clinical trials IFN α , usually combined with RA, was given to patients with squamous cell carcinoma (SCC) of uterine cervix; such an approach is of special interest, since both agents have been shown to suppress the growth of human papilloma virus type 16 (HPV-16)(109), which is related to cervical carcinoma. Moreover, IFN might correct the RT-due, long-lasting depression of lymphocytes (110,111). However, the results obtained in these trials are very heterogeneous. The high OR rates of 50% and 42% reported by Lippman et al. (112,113) were not confirmed in later studies (114,115).

The Lippman's group also reported very high (68%) objective response of patients with inoperable SCC of the skin; again, IFN α was combined with retinoic acid (116). The high response rates seen in these initial trials were not obtained in other squamous tumors (head and neck, oesophagus), in which this regimen (117-119), or the combination of IFN with IL-2 (120,121), was used. Further studies integrating such therapy with other treatment modalities are warranted in cervix and skin cancers.

Interferons were also tried as local-intravesical therapy in superficial bladder cancer. The average OR rate was 40% (122,123). Intravesical instillation of IFN lowers the relapse rate from 70-80%, seen after surgery alone, to 30-50%. It is noteworthy that this treatment has few and usually mild side effects, which is in contrast to the routine local BCG therapy; however, the latter has significantly higher response rates (60-70%). Because of that, an ongoing multi-center trial (123) uses low doses of both agents in the therapy of superficial bladder cancer.

The local (intra- and perilesional) or systemic IFN therapy was tried, with some success, in patients with various other solid tumors: hondrosarcoma (124), malignant pleural mesothelioma (125,126), glioma (127), prostatic cancer (128) and AIDS-related Kaposi's sarkoma (129,130). This latter entity is of special interest, since it is the most common complication of HIV infection and AIDS; therefore, all biological activities of IFN (antiviral, antitumor and immunological), may be of relevance. It is agreed that IFN monotherapy may be effective in a proportion of patients (those with CD4 cell number > 150 cells/mm³)(131). However, the doses necessary to achieve a significant antitumor effect are often poorly tolerated. The therapy with IFN and zidovudine (132-135) resulted in tumor regression in a substantial percentage

of patients, but was usually associated with dose-limited toxicity. Generally, this disease remains a challenging problem; larger studies using IFN combined with antiretroviral treatment and chemotherapy are warranted.

CONCLUSIONS AND FUTURE DIRECTIONS

Although IFNs have been tried in the therapy of solid malignancies other than RCC and MM for several years, it can be stated presently that this approach is still in experimental area. Despite the wide variety of clinical trials using IFNs either as monotherapy or in combination with other BRMs, no clear-cut enhancement of therapeutic efficacy of standard treatment has been substantiated. However, it seems that some combinations, such as IFN and hormone therapy for breast cancer, and IFN+retinoic acid in squamous cell carcinomas, deserve further investigation. The disappointing results of IFN therapy may be primarily due to its use mainly in advanced cancer. As it has been known for several years, the biotherapy might be more effective in the early stages of tumor development. Therefore, the optimal effects of IFN therapy may be expected during the early evolution of cancer (136), and in pre-malignant lesions as well (137,138). The kinetics of tumor cell populations influence the expression of specific receptors, which is a common denominator of the action of BRMs. Thus, the response of human tumors to IFN strongly depends on the tumor cell growth kinetics (136,137): the stationary cell populations are killed, whereas the fast growing ones are only reversibly inhibited. This may be the reason for different sensitivity of primary tumors and metastases (139).

In future investigations of therapeutic potential of IFNs in solid malignancies, the key question to be addressed is the better understanding of biological mechanisms of specific tumor sensitivity, i.e., why some tumors of the same histologic type are responsive and some are not. Furthermore, the mechanisms of resistance of sensitive tumor types also have to be resolved. Such research may lead to better defining the biological features related to tumor responsiveness (140, 141).

In addition, several other points remain to be elucidated: the relative contribution of antiviral, antitumor and immunologic effects of IFNs in exerting the beneficial effect in some malignant lesions, the defining predictive factors for clinical response and the optimal therapeutic schedule, the reduction of toxicity, the treatment duration, new indications and new drug combinations - all are expected to be resolved in near future.

Acknowledgements

We thank Mr Novak Vuletić for his help in preparing the manuscript.

REFERENCES

1. Stanojević-Bakić N, Vučković-Dekić Lj. Interferons in therapy of solid tumors. Part I. Interferons and renal cell carcinoma. *Arch Oncol* 1999;7:75-80.
2. Vučković-Dekić Lj, Stanojević-Bakić N. Interferons in the therapy of solid tumors. Part II. Interferons and malignant melanoma. *Arch Oncol* 1999;7:176.
3. Kjaer M. Combining 5-fluorouracil with interferon- α in the treatment of advanced colorectal cancer: optimism followed by disappointment. *Anti-Cancer Drugs* 1996;7:35-42.
4. Lillis PK, Brown TD, Beougher K, Koeller J, Marcus SG, Von Hoff DD. Phase II trial of recombinant beta interferon in advanced colorectal cancer. *Cancer Treat Rep* 1987;71:965-7.
5. O'Connell MJ, Ritts RA, Moertel CG. Recombinant interferon-g lacks activity against metastatic colorectal cancer but increases serum levels of CA-19-9. *Cancer* 1989;63:1998-2004.
6. Bendtzek K, Hansen MB, Diamant M, Ross C, Svenson M. Naturally occurring autoantibodies to interleukin 1a, interleukin 6, interleukin 10 and interferon α . *J Interferon Res* 1994;14:157-8.
7. Karmaniolas KD, Papalampros TS, Papavassiliou D, Liatis STH, Kalantzis N. Detection of interferon inhibitors or antagonists in gastrointestinal malignancies. *Hepato-Gastroenterology* 1998;45:2244-7.
8. Schwartz EL, Hoffman M, O'Connor CJ. Stimulation of 5-fluorouracil metabolic activation by interferon- α in human colon carcinoma cells. *Biochem Biophys Res Commun* 1992;182:1232-9.
9. Huschart T, Medenica R, Hankenson R, Corbitt W. Response to interferon therapy is dependent on interferon inhibitor factor levels. *J Interferon Res* 1994;14:116-20.
10. Clark RI, Slevin ML, Reznick RH. Two randomised phase II trials of intermittent intravenous versus subcutaneous alpha-2-interferon alone (trial 1) and in combination with 5-fluorouracil (trial 2) in advanced cancer. *Int J Colon Dis* 1987;2:262-9.
11. Wadler S, Wiernik PH. Clinical up-date on the role of fluorouracil and recombinant interferon alpha-2a in the treatment of colorectal carcinoma. *Semin Oncol* 1990;17 (suppl 1):16-21.
12. Pazdur R, Ajani JA, Patt YZ. Phase II study of fluorouracil and recombinant interferon alpha-2a in previously untreated advanced colorectal carcinoma. *J Clin Oncol* 1990;8:2027-31.
13. Kemeny N, Younes A, Seiter K, Kelsen D, Sammarco P, Adams L et al. Interferon alpha-2a and 5-fluorouracil for advanced colorectal carcinoma: assessment of activity and toxicity. *Cancer* 1990; 66:2470-5.
14. Fornasiero A, Daniele O, Ghiotto C. Alpha-2a interferon and 5-fluorouracil in advanced colorectal cancer. *Tumori* 1990;76:385-8.
15. Wadler S, Lembersky B, Atkins M, Kirkwood J, Petrelli N. Phase II trial of 5-fluorouracil and recombinant interferon alpha-2a in patients with advanced colorectal carcinoma: an Eastern Cooperative Oncology Group study. *J Clin Oncol* 1991;9:1806-10.
16. Meadows LM, Walther P, Ozer H. a-Interferon and 5-fluorouracil: possible mechanisms of antitumor action. *Semin Oncol* 1991;18(Suppl 7):71-6.
17. Weh HJ, Platz D, Braumann D. Phase II trial of 5-fluorouracil and recombinant interferon alpha-2B in metastatic

colorectal carcinoma. *Eur J Cancer* 1992;28A:1820-3.

18. Rubio D, Jimeno J, Camps C. Treatment of advanced colorectal cancer with recombinant interferon alpha and fluorouracil: activity in liver metastases. *Cancer Invest* 1992;10:259-64.
19. Pazdur R, Ajani JA, Patt YZ, Gomez J, Bready B, Levin B. Phase II evaluation of recombinant alpha-2a-interferon and continuous infusion fluorouracil in previously untreated metastatic colorectal adenocarcinoma. *Cancer* 1993;71:1214-8.
20. John WJ, Neefe JR, MacDonald JS, Cantrell J, Smith M. 5-fluorouracil and interferon-alpha-2a in advanced colorectal cancer. Results of two treatment schedules. *Cancer* 1993;72:3191-5.
21. Findlay M, Hill A, Cunningham D. Protracted venous infusion 5-fluorouracil and interferon- α in advanced and refractory colorectal cancer. *Ann Oncol* 1994;5:239-43.
22. CORFU-A Study Group. Phase II randomized study of two fluorouracil combinations with either interferon-alpha-2a or leucovorin for advanced colorectal cancer. *J Clin Oncol* 1995;13:921-8.
23. Hill MH, Norman A, Cunningham D, Findlay M, Watson M, Nicolson V et al. Royal Marsden phase II trial of fluorouracil with or without interferon alpha-2b in advanced colorectal cancer. *J Clin Oncol* 1995;13:1297-1302.
24. Piga A, Cascinu S, Latini L, Marcellini M, Bavosi M, Acito L et al. A phase II randomised trial of 5-fluorouracil with or without interferon alpha-2a in advanced colorectal cancer. *Br J Cancer* 1996;74:971-4.
25. Dufour P, Husseini F, Dreyfus B, Cure H, Martin C, Puvost G et al. 5-Fluorouracil versus 5-fluorouracil + a-interferon as treatment of metastatic colorectal carcinoma. A randomized study. *Ann Oncol* 1996;7:575-9.
26. Patt YZ, Hoque A, Lozano R, Pazdur R, Chase J, Carrasco H et al. Phase II trial of hepatic arterial infusion of fluorouracil and recombinant human interferon alpha-2b for liver metastases of colorectal cancer refractory to systemic fluorouracil and leucovorin. *J Clin Oncol* 1997;15:1432-8.
27. Perez JE, Lacava JA, Dominguez ME, Rodriguez R, Barbieri MR, Romero-Acuna LA et al. Biomodulation with sequential intravenous IFN-alpha2b and 5-fluorouracil as second-line treatment in patients with advanced colorectal cancer. *J Interferon Cytokine Res* 1998; 18:565-9.
28. Kim J, Zhi J, Satoh H, Koss-Twardy SG, Passe SM, Patewl IH et al. Pharmacokinetics of recombinant human interferon-alpha 2a combined with 5-fluorouracil in patients with advanced colorectal carcinoma. *Anti-Cancer Drugs* 1998;9:689-96.
29. Kohne CH, Schoffski P, Wilke H, Kaufer C, Andreesen R, Ohl U et al. Effective biomodulation by leucovorin of high-dose infusion fluorouracil given as a weekly 24-hour infusion: results of a randomized trial in patients with advanced colorectal cancer. *J Clin Oncol* 1998;16:418-26.
30. Joffe JK, Perren TJ, Bradley C, Primrose J, Hallam S, Ward U et al. A phase II study of recombinant interferon-beta (r-hIFN- β 1a) in combination with 5-fluorouracil (5-FU) in the treatment of patients with advanced colorectal carcinoma. *Br J Cancer* 1997;75:423-6.
31. Wadler S, Haynes H, Rosenblit A, Hu X, Kaleya R, Wiernik PH. Sequential phase II trials of fluorouracil and interferon beta ser with or without sargramostim in patients with advanced colorectal carcinoma. *Cancer J Sci Am* 1998;4:331-7.
32. Villar-Grimalt A, Candel MT, Massuti B, Lizon J, Sanchez B, Frau A et al. A randomized phase II trial of 5-fluorouracil with or without human interferon-beta for advanced colorectal cancer. *Br J Cancer* 1999;80:786-91.
33. Raderer M, Scheithauer W. Treatment of advanced colorectal cancer with fluorouracil and interferon- α : an overview of clinical trials. *Eur J Cancer* 1995;31A:1002-8.



34. Kase S, Kubota T, Watanabe M, Furukawa T, Tanino H, Ishibiki K et al. Interferon beta increases antitumor activity of 5-fluorouracil against human colon carcinoma cell lines in vitro and in vivo. *Anticancer Res* 1993;13:69-74.
35. Sobrero A, Nobile MT, Guglielmi A, Mori A, Aschele C, Bolli E et al. Phase II study of 5-fluorouracil plus leucovorin and interferon alpha-2b in advanced colorectal cancer. *Eur J Cancer* 1992;28A:850-2.
36. Grem JL, Jordan E, Robson ME, Binder RA, Hamilton JM, Steinberg SM et al. Phase II study of fluorouracil, leucovorin and interferon-alfa-2a in metastatic colorectal carcinoma. *J Clin Oncol* 1993;11:1737-45.
37. Punt CJ, Burhouts JT, Croles JJ, van Liessum PA, de Mulder PH, Kamm Y. Continuous infusion of high-dose 5-fluorouracil in combination with leucovorin and recombinant interferon-alpha-2b in patients with advanced colorectal cancer. A multicenter phase II study. *Cancer* 1993;72:2107-11.
38. Pazdur R, Roach R, Braud E. Phase II study of folinic acid, 5-fluorouracil, and interferon alfa-2a for patients with advanced colorectal carcinoma. *Proc Am Soc Clin Oncol* 1994;13:224-7.
39. Kosmidis PA, Tsavaris N, Skarlos D, Theocharis D, Samantas E, Pavlidis N et al. Fluorouracil and leucovorin with or without interferon alfa-2b in advanced colorectal cancer: Analysis of a prospective randomized phase III trial. Hellenic Cooperative Oncology Group. *J Clin Oncol* 1996;14:2682-7.
40. Tournigand C, Louvet C, de Gramont A, Lucchi E, Seitz JF, Mal F et al. Bimonthly high dose leucovorin and 5-fluorouracil 48-hour infusion with interferon-alpha-2a in patients with advanced colorectal carcinoma. Groupe d'Etude et de Recherche sur les Cancers de l'Ovaire et Digestifs (GERCOD). *Cancer* 1997;79:1094-9.
41. Kohne CH, Wilke H, Hiddemann W, Bokemeyer C, Lohrmann HP, Bodenstein H et al. Phase II evaluation of 5-fluorouracil+folinic acid and alpha 2b-interferon in metastatic colorectal cancer. *Oncology* 1997;54:96-101.
42. Kohne CH, Schoffski P, Wilke H, Kaufner C, Andreesen R, Ohl U et al. Effective biomodulation by leucovorin of high-dose infusion fluorouracil given as a weekly 24-hour infusion. Results of randomized trial in patients with advanced colorectal cancer. *J Clin Oncol* 1998;16:418-26.
43. Hausmaninger H, Moser R, Samonigg H, Mlineritsch B, Schmidt H, Pecherstorfer M et al. Biochemical modulation of 5-fluorouracil by leucovorin with or without interferon- α -2c in patients with advanced colorectal cancer: final results of a randomised phase III study. *Eur J Cancer* 1999;35:380-5.
44. Ajani JA, Abbruzzese JL, Markowitz AB, Patt YZ, Daughtery K. Phase II study of etoposide and alpha-interferon in patients with advanced measurable colorectal cancer. *Invest New Drugs* 1993;11:67-9.
45. Royce ME, McGarry W, Bready B, Dakhil SR, Bett RJ, Goodwin JW et al. Sequential biochemical modulation of fluorouracil with folinic acid, N-phosphonacetyl-L-aspartic acid and interferon alfa-2a in advanced colorectal cancer. *J Clin Oncol* 1999;17:3276-82.
46. Ridolfi R, Maltoni R, Riccoban A, Flamini E, Fedriga R, Milandri C et al. A phase II study of advanced colorectal cancer patients treated with combination 5-fluorouracil plus leucovorin and subcutaneous interleukin-2 plus alpha interferon. *J Chemother* 1994;6:265-71.
47. Atzpodiën J, Kirchner H, Hanninen EL, Menzel T, Deckert M, Franzke A et al. Treatment of metastatic colorectal cancer patients with 5-fluorouracil in combination with recombinant subcutaneous human interleukin-2 and alpha-interferon. *Oncology* 1994;51:273-5.
48. Goey SH, Gratamo JW, Primarose JH, Ward U, Mertelmann RH, Osterwalder B. Interleukin 2 and interferon alpha-2a do not improve anti-tumour activity of 5-fluorouracil in advanced colorectal cancer. *Br J Cancer* 1996;74:2018-23.
49. Grem JL, McAtee N, Murphy RF, Balis FM, Cullen E, Chen AP et al. A pilot study of gamma-1b-interferon in combination with fluorouracil, leucovorin, and alpha-2a-interferon. *Clin Cancer Res* 1997;3:1125-34.
50. Wadler S, Bajaj R, Neuberg D, Agarwal V, Haynes H, Benson AB. Prognostic implications of c-Ki-ras2 mutations in patients with advanced colorectal cancer treated with 5-fluorouracil and interferon: a study of the Eastern Cooperative Oncology Group (EST 2292). *Cancer J Sci Amer* 1997;3:284-8.
51. Tsavaris N, Baxevas C, Kosmidis P, Papamichael M. The prognostic significance of immune changes in patients with renal cancer, melanoma and colorectal cancer, treated with interferon alpha 2b. *Cancer Immunol Immunother* 1996;43:94-102.
52. Meredith RF, Khazaeli MB, Plott WE, Guizzh WE, Lui T, Schlom J et al. Phase II study of dual 131I-labeled monoclonal antibody therapy with interferon in patients with metastatic colorectal cancer. *Clin Cancer Res* 1996;2:1811-8.
53. Roselli M, Guadagni F, Buonomo O, Belardi A, Vittomini V, Mariani-Constantini R et al. Systemic administration of recombinant interferon alfa in carcinoma patients upregulates the expression of the carcinoma-associated antigens tumor-associated glycoprotein-72 and carcinoembryonic antigen. *J Clin Oncol* 1996;14:2031-42.
54. John WL, Foon KA. Clinical applications of interferon in other tumors. In: DeVita, Hellman S, Rosenberg SA, eds. *Biologic therapy of cancer: principles and practice*. Philadelphia: Lippincott-Raven, 1995:427-33.
55. Schiller JH, Storer B, Dreicer R, Rosenquist D, Frontiera M, Carbone PP. Randomized phase II-III trial of combination beta and gamma interferons and etoposide and cisplatin in inoperable non-small cell cancer of the lung. *J Natl Cancer Inst* 1989;81:1739-43.
56. Bowman A, Fergusson RJ, Allan SG. Potentiation of cisplatin by alpha-interferon in advanced non-small cell lung cancer (NSCLC): a phase II study. *Ann Oncol* 1990;1:351-3.
57. Rosell R, Carles J, Ariza A, Moreno I, Ribelles N, Solano V et al. A phase II study of days 1 and 8 cisplatin and recombinant alpha-2B interferon in advanced non-small cell lung cancer. *Cancer* 1991;15:2448-53.
58. Lind MJ, Gomm S, Simmonds AP, Ashcroft L, Kamthan A, Gurney H et al. A phase II study of ifosfamide and alpha-2b interferon in advanced non-small-cell lung cancer. *Cancer Chemother Pharmacol* 1991;28:142-4.
59. Ardizzoni A, Salvati F, Rosso R. Combination of chemotherapy and recombinant alpha-interferon in advanced non-small cell lung cancer: multicenter randomized FONICAP trial report. *Cancer* 1993;72:2929-35.
60. Halme M, Maasilta PK, Pyrhonen SO, Mattson KV. Interferons combined with chemotherapy in the treatment of stage III-IV non-small cell lung cancer - a randomised study. *Eur J Cancer* 1994;30A:11-5.
61. Quan DYW, Madajewicz S, Smith MR. Alpha interferon, leucovorin, and 5-fluorouracil (ALF) in advanced cancer: results of a dose-finding study and evidence of activity in non-small cell lung cancer. *Cancer Invest* 1994;12:367-74.
62. Quan DYW, Casal R, Rosenfeld M, Walker RP. Alpha interferon-2b, leucovorin, and 5-fluorouracil (ALF) in non-small cell lung cancer. *Cancer Biother Radiopharm* 1996;11:229-34.
63. Prior C, Oroszy S, Oberaigner W, Ambrosch G, Mohn-Staudner A, Pfeifer W et al. Advanced non-small cell lung cancer: adjunctive interferon γ in induction and maintenance therapy. *J Cancer Res Clin Oncol* 1999;125:42-6.
64. Maasilta P, Holsti LR, Halme M, Kivisaari L, Cantell K, Maltson K. Natural alpha-interferon in combination with hyperfractionated radiotherapy in the treatment of non-small cell lung cancer. *Int J Radiat Oncol Biol Phys* 1992;23:863-8.
65. Mc Donald S, Chang AY, Rubin P, Wallenberg J, Kim IS, Sobel S et al. Combined betaseron R (recombinant human interferon beta) and radiation for inoperable non-small cell lung cancer. *Int J Radiat Oncol Biol Phys* 1993;20:753-6.
66. Shaw EG, Deming DL, Creagan ET, Nair S, Su JQ, Levitt R et al. Pilot study of human recombinant interferon gamma and accelerated hyperfractionated thoracic radiation therapy in patients with unresectable stage IIIA/B non-small cell lung cancer. *Int J Radiat Oncol Biol Phys* 1995;31:827-31.
67. Krigel R, Padovic K, Weiner L. Phase I study of sequential recombinant tumor necrosis factor (rTNF) and recombinant interleukin-2 (rIL-2). *Proc Am Assoc Cancer Res* 1991;32:268-71.
68. Rinaldi DA, Lippman SM, Burris HA, Chon C, Von Hoff D, Hong WK. Phase II study of 13-cis-retinoic acid and interferon-alpha-2a in patients with advanced squamous cell lung cancer. *Anti-Cancer Drugs* 1993;4:33-6.
69. Arnold A, Ayong J, Douglas L, Hoogendoorn P, Skingley L, Gelmon K et al. Phase II trial of 13-cis-retinoic acid plus interferon α in non-small cell lung cancer. *J Natl Cancer Inst* 1994;86:306-9.
70. Athanasiadis I, Kies MS, Miller M, Ganzenko N, Joob A, Marymont M et al. Phase II study of all-trans-retinoic acid and α -interferon in patients with advanced non-small cell lung cancer. *Clin Cancer Res* 1995;1:973-9.
71. Garaci E, Lopez M, Bonsignore G. Sequential chemioimmunotherapy for advanced non-small cell lung cancer using cisplatin, etoposide, thymosin- α_1 and interferon- α_2 . *Eur J Cancer* 1995;31A:2403-5.
72. Salvati F, Rasi G, Portalone L, Antili A, Garaci E. Combined treatment with thymosin-alpha1 and low-dose interferon-alpha after ifosfamide in non-small cell lung cancer: a phase II controlled trial. *Anticancer Res* 1996;16:1001-4.
73. Kohne-Wompner CH, Koschel G, Pawel JV, Wilke H, Vallee D, Bremer K et al. Small-cell lung cancer (SCLC) limited disease (LD): carboplatin (C), etoposide (E), vincristine (V) chemotherapy (CT) and radiotherapy (RT) followed by α 2b interferon (IFN) maintenance vs observation for patients (pts) with response after CT/RT. *Ann Oncol* 1992;(53):40.
74. Mattson K, Niiranen A, Pyrhonen S. Natural interferon alfa as maintenance therapy for small cell lung cancer. *Eur J Cancer* 1992;28:1387-91.
75. Mattson K, Niiranen A, Ruotsalainen T, Maasilta P, Halme M, Pyrhonen S et al. Interferon maintenance therapy for small-cell lung cancer: improvement in long term survival. *J Interferon Cytokine Res* 1997;17:103-5.
76. Kelly K, Punn P, Hazuka M. The role of alpha interferon maintenance in patients with limited stage SCLC responding to concurrent chemoradiation. A SWOG study. *Proc Am Soc Clin Oncol* 1993;12:330 (Abstr. 1099).
77. Glisson BS, Palmer JL, Shin DM, Wester M, Markowitz AB. Phase II trial of interferon alpha (IFN) plus etoposide/cisplatin (EP) induction and IFN/Megau (M) maintenance in extensive small cell lung cancer (ESCLC). *Proc Am Soc Clin Oncol* 1993;12:354.
78. Jett RJ, Maksymiuk WA, Su QJ, Mailliard JA, Krook EJ, Tschetter KL et al. Phase III trial of recombinant interferon gamma complete responders with small-cell lung cancer. *J Clin Oncol* 1994;12:2321-6.
79. Bitran DJ, Green M, Perry M, Hollis RD, Herndon EJ. A phase II study of recombinant interferon-gamma following combination chemotherapy for patients with extensive small cell lung cancer. *Am J Clin Oncol* 1995;18:67-70.
80. vanZandwijk N, Groen HJM, Postmus PE, Burghouts JTW, Velde GPM, Ardizzoni A et al. Role of recombinant interferon-gamma maintenance in responding patients with

- small cell lung cancer. A randomised phase III study of the EORTC Lung Cancer Cooperative Group. *Eur J Cancer* 1997;33:1759-66.
81. Tummarello D, Mari D, Graciano F, Isidori P, Cetto G, Pasini F et al. A randomized controlled phase III study of cyclophosphamide, doxorubicin, and vincristine with etoposide (CAV-E) or teniposide (CAV-T), followed by recombinant interferon- α maintenance therapy or observation, in small cell lung carcinoma patients with complete responses. *Cancer* 1997;80:2222-9.
82. Zarogoulidis K, Ziogas E, Papagiannis A, Charitopoulos K, Dimitriadis K, Economides D et al. Interferon alpha-2a and combined chemotherapy as first line treatment in SCLC patients: a randomised trial. *Lung Cancer* 1996;15:197-205.
83. Prior C, Oroszy S, Oberaigner W, Schenk E, Kummer F, Aigner K et al. Adjunctive interferon-alpha-2c in stage IIIB/IV small-cell lung cancer: a phase III trial. *Eur Respir J* 1997;10:392-6.
84. Hadden JW. The immunology and immunotherapy of breast cancer: an update. *Int J Immunopharmacol* 1999;21:79-101.
85. Laszlo J, Hood L, Cox E, Goodwin B. A randomized trial of low doses of alpha interferon in patients with breast cancer. *J Biol Res Modif* 1986;5:206-10.
86. Fentiman SI, Balkwill RF, Cuzick J, Hayward LJ, Rubens DR. A trial of human alpha interferon as an adjuvant agent in breast cancer after loco-regional recurrence. *Eur J Surg Oncol* 1987;13:425-8.
87. Stanojević-Bakić N, Vučković-Dekić Lj, Inić M, Tomin R. Effect of interferon treatment on general immune competence in breast cancer patients. *Glas de l'Academie Serbe des Sciences et des Arts* 1994;44:71-82.
88. Walters RS, Theriault RL, Booser DJ, Espazza L, Hortobagyi GN. Phase II study of recombinant alpha-interferon (rIFN alpha) and continuous-infusion 5-fluorouracil in metastatic breast cancer. *J Immunother Emphasis Tumor Immunol* 1995;18:185-7.
89. Oldham RK, Blumenschein G, Schwartzberg L. Continuous biotherapy utilizing interleukin-2 and alpha interferon in patients with advanced cancer: a National Biotherapy Study Group trial. *Mol Biother* 1992;4:March.
90. Walters RS, Theriault RL, Holmes FA, Espazza L, Hortobagyi GN. Phase II study of recombinant alpha-interferon and recombinant interleukin-2 in metastatic breast cancer. *J Immunother Emphasis Tumor Immunol* 1994;16:303-5.
91. Tonini G, Nunziata G, Prete SP, Peponi R, Turriziani M, Masci G et al. Adjuvant treatment of breast cancer: a pilot immunotherapy study with CMF, interleukin-2 and interferon alpha. *Cancer Immunol Immunother* 1998;47:157-66.
92. Ozzello L, Habif DV, DeRosa CM, Cantell K. Cellular events accompanying regression of skin recurrences of breast carcinomas treated with intralesional injections of natural interferons α and γ . *Cancer Res* 1992;52:4571-81.
93. Habif DV, Ozzello L, DeRosa CM, Cantell K, Lattes R. Regression of skin recurrences of breast carcinomas treated with intralesional injections of natural interferons alpha and gamma. *Cancer Invest* 1995;13:165-72.
94. Munno I, Marinario M, Gesario A, Cannuscio B, Michel Y, Paulling E. Immunomodulatory effects of alpha interferon and thymostimulin in patients with neoplasias. *Clin Diagn Lab Immunol* 1995;July:503-5.
95. Bezwoda WR, Meyer K. Effect of alpha-interferon, 17 β -oestradiol and tamoxifen on oestrogen receptor concentration and cell cycle kinetics of MFC7 cells. *Cancer Res* 1990;50:5387-91.
96. Coradini D, Biffi A, Pirronello E, DiFronzo G. Tamoxifen and beta-interferon: effect of simultaneous or sequential treatment on breast cancer cell lines. *Anticancer Res* 1995;15:315-9.
97. DiMartino L, Demontis B, Saccani Iotti G, Murenu G. *In vivo* effect induced by interferon beta on steroid receptor status, cell kinetics and DNA ploidy in operable breast cancer patients. *Anticancer Res* 1995;15:537-41.
98. Buzzi F, Burgia M, Rossi G, Giustini L, Scoconi G, Sica G. Combination of beta-interferon and tamoxifen as a new way to overcome clinical resistance to tamoxifen in advanced breast cancer. *Anticancer Res* 1992;12:869-71.
99. Seymour L, Bezwoda WR. Interferon plus tamoxifen treatment for advanced breast cancer: in vivo biologic effects of two growth modulators. *Br J Cancer* 1993;68:352-6.
100. Buzzi E, Burgia M, Trippa F, Rossi G, Trivisonne R, Guistini L et al. Natural interferon-beta and tamoxifen in hormone-resistant patients with advanced cancer. *Anticancer Res* 1995;15:2187-90.
101. Repetto L, Giannesi PG, Campora E, Pronzato P, Vignani A, Naso C et al. Tamoxifen and interferon-beta for the treatment of metastatic breast cancer. *Breast Cancer Res Treat* 1996;39:235-8.
102. Kornek G, Reiner A, Sagaster P, Stierer M, Mayer A, Ludwig H. Effect of interferon alpha-2a on hormone receptor status in patients with advanced breast cancer. *Cancer Invest* 1999;117:189-94.
103. Barak V, Kalickman I, Nisman B, Farbstein H, Fridlender ZG, Baicler L et al. Changes in cytokine production of breast cancer patients treated with interferons. *Cytokine* 1998;10:977-83.
104. Lindner DJ, Borden EC, Kalvakolanu DV. Synergistic antitumor effects of a combination of interferons and retinoic acid on human tumor cells *in vitro* and *in vivo*. *Clin Cancer Res* 1997;3:931-7.
105. Lama G, Angeluca C, Recchia F, Sica G. Combined effect of 13-cis-retinoic acid, tamoxifen and interferon on the growth of human breast cancer. *Cancer Lett* 1996;149:203-8.
106. Recchia F, Sica G, DeFilippis S, Discepoli S, Rea S, Torchio P et al. Interferon-beta, retinoids and tamoxifen in the treatment of metastatic breast cancer: a phase II study. *J Interferon Cytokine Res* 1995;15:605-10.
107. Recchia F, Rea S, DeFilippis S, Rosselli M, Corrao G, Gulino A, Sica G. Beta-interferon and tamoxifen combination in advanced breast cancer. *Clin Ther* 1998;149:203-8.
108. Recchia F, Frati L, Rea S, Torchio P, Sica G. Minimal residual disease in metastatic breast cancer: treatment with IFN-beta, retinoids and tamoxifen. *J Interferon Cytokine Res* 1998;18:41-7.
109. Agarwal C, Hembree JR, Rorke EA. Interferon and retinoic acid suppress the growth of human papilloma virus type 16 immortalized cervical epithelial cells, only interferon suppresses the level of human papilloma virus transforming oncogenes. *Cancer Res* 1994;54:2108-12.
110. Angioli R, Sevin BU, Perras JP, Untch M, Hightower R, Nguyen HN et al. Rationale of combining radiation and interferon for the treatment of cervical cancer. *Oncology* 1992;49:445-9.
111. Vučković-Dekić Lj, Spremo B, Stanojević-Bakić N, Garžičić B, Durbaba M. Immunosuppressive and cytogenetic effects of pelvic irradiation on the peripheral lymphocytes of patients with cervical cancer. *Arch Immunol Ther Exp* 1994;42:63-6.
112. Lippman SM, Kavanagh JJ, Paredes-Espinoza M. 13-cis-retinoic acid+interferon α -2a in locally advanced squamous cell carcinoma of the cervix. *J Natl Cancer Inst* 1993;85:499-500.
113. Lippman SM, Kavanagh JJ, Paredes M. 13-cis-retinoic acid (13 cRA), interferon α -2a (IFN- α a) and radiotherapy for locally advanced cancer of the cervix. *Proc Am Soc Clin Oncol* 1993; 12:257 (abstr.).
114. Murad AM, Oliveira M, Saldanha TM. Phase II trial of isotretinoin and interferon α -2a in the treatment of advanced recurrent cervical carcinoma. *Int J Gynecol Cancer* 1994;4:414-8.
115. Hallum AV, Alberts DS, Lippman SM. Phase II study of 13-cis retinoic acid + interferon-alpha 2a in heavily pretreated squamous carcinoma of the cervix. *Gynecol Oncol* 1995;56:382-6.
116. Lippman SM, Parkinson DR, Itri LM, Weber RS. 13-cis retinoic acid and interferon α -2a: effective combination therapy for advanced squamous cell carcinoma of the skin. *J Natl Cancer Inst* 1992;84:235-41.
117. Temek KB, Liebmann EJ, Theodossiou C, Steinberg MS, Cook AJ, Metz CD et al. Phase II trial of 5-fluorouracil, leucovorin, interferon- α -2a, and cisplatin as neoadjuvant chemotherapy for locally advanced oesophageal carcinoma. *Cancer* 1996;77:2432-9.
118. Kok TC, van der Gaast, Splinter TAW. 12-cis-retinoic acid and alpha-interferon in advanced squamous cell cancer of the oesophagus. *Eur J Cancer* 1997;33:165-6.
119. Vlock DR, Andersen J, Kalish LA, Johnson JT, Kirkwood JM, Whiteside T et al. Phase II trial of interferon- α in locally recurrent or metastatic squamous cell carcinoma of the head and neck: immunological and clinical correlates. *J Immunother* 1997;19:433-42.
120. Schantz SP, Dimetry I, Lippman SM. A phase II study of interleukin-2 and interferon-alpha in head and neck cancer. *Invest New Drugs* 1992;10:217-33.
121. Urba SG, Forastiere AA, Wolf GT. Intensive recombinant interleukin-2 and alpha-interferon therapy in patients with advanced head and neck squamous carcinoma. *Cancer* 1993;71:2326-31.
122. Tanneberger S, Hrelia P. Interferons in precancer and cancer prevention: where are we? *J Interferon Cytokine Res* 1996;16:339-46.
123. Gan YH, Mahendran R, James K, Lawrence C, Esuvaranathan K. Evaluation of lymphocytic responses after treatment with Bacillus Calmette-Guerin and interferon- α 2b for superficial bladder cancer. *Clin Immunol* 1999;90:230-7.
124. Baltić VV. Intratumor immunotherapy chondrosarcoma pelvis with recombinant interferon alpha 2a. *Arch Oncol* 1993;1:47-9.
125. Purohit A, Moreau L, Dietemann A, Seibert R, Pauli G, Wihlm JM et al. Weekly systemic combination of cisplatin and interferon α 2a in diffuse malignant pleural mesothelioma. *Lung Cancer* 1998;22:119-25.
126. Trandafir L, Ruffie P, Borel C, Monnet I, Soulie P, Adams D et al. Higher doses of α -interferon do not increase the activity of the weekly cisplatin-interferon combination in advanced malignant mesothelioma. *Eur J Cancer* 1997;11:1900-2.
127. Buckner JC, Brown LD, Kugler JW, Cascino TL, Krook JE, Mailliard JA et al. Phase II evaluation of recombinant interferon alpha and BCNU in recurrent glioma. *J Neurosurg* 1995;82:430-5.
128. Morales A, Johnson B, Emerson L, Heaton JWP. Intralesional administration of biological response modifiers in the treatment of localized cancer of the prostate: a feasibility study. *Urology* 1997;50:495-502.
129. Krown S. Approaches to interferon combination therapy in the treatment of AIDS. *Semin Oncol* 1990;17(Suppl 1):11-5.
130. Salma S, Baltić V, Todorovic V. Clinical effects of interferon- α -2a treatment of Kaposi's sarcoma. *Arch Oncol* 1994;2:223.
131. Sneller MC. Consensus symposium on combined antiviral therapy: overview of interferon and IL-2 combinations for the treatment of HIV infection. *Antiviral Res* 1996;29:15-9.



132. Fischl MA, Finkelstein DM, He W, Powderly WG, Triozzi PL, Steigbigel RT. A phase II study of recombinant human interferon-alpha 2a and zidovudine in patients with AIDS-related Kaposi's sarcoma. AIDS Clinical Trial Group. J Acquir Immune Defic Syndr Hum Retrovirol 1996;11:379-84.
133. Shepherd FA, Beaulieu R, Gelmon K, Thuot CA, Sawka C, Read S et al. Prospective randomized trial of two dose levels of interferon alfa with zidovudine for the treatment of Kaposi's sarcoma associated with human immunodeficiency virus infection: a Canadian HIV Clinical Trials Network study. J Clin Oncol 1998;16:1736-42.
134. Miles S, Levine A, Feldstein M, Carden J, Cabriallas S, Marcus S et al. Open-label phase I study of combination therapy with zidovudine and interferon-beta in patients with AIDS-related Kaposi's sarcoma: AIDS Clinical Trials Group Protocol 057. Cytokines Cell Mol Ther 1998;4:17-23.
135. Aversa SM, Cattelan AM, Salvagno L, Meneghetti F, Francavilla E, Sattin L et al. Chemo-immunotherapy of advanced AIDS-related Kaposi's sarcoma. Tumori 1999;85:54-9.
136. Gutterman JU. Cytokine therapeutics: lessons from interferon- α . Proc Natl Acad Sci USA 1994;91:1198-205.
137. Tanneberger S, Hrelia P. Interferons in precancer and cancer prevention: where are we? J Interferon Cytokine Res 1996;16:339-46.
138. Kuwana K, Ichida T, Kamimura T, Ohkoshi S, Ogata N, Harada T et al. Risk factors and the effect of interferon therapy in the development of hepatocellular carcinoma: A multivariate analysis in 343 patients. J Gastroenterol Hepatol 1997;12:149-55.
139. Tanneberger S, Pannuti F. Disillusionments and hopes in the field of biological response modifiers. Anticancer Res 1993;13:185-92.
140. Eisenhauer EA, Lippman SN, Kavanagh JJ, Parades-Espinoza M, Arnold A, Hong WK et al. Combination 13-cis-retinoic acid and interferon alpha-2a in the therapy of solid tumors. Leukemia 1994;8(Suppl.3):S38-41.
141. Witt PL, Lindner DJ, D'Cunha , Borden CE. Pharmacology of interferons: induced proteins, cell activation, and antitumor activity. In: Chabner BA, Longo DL, eds. Cancer Chemotherapy and Biotherapy, second edition. Philadelphia: Lippincott-Raven Publishers, 1996:585-607.



The next ESMO congress will be held in Hamburg from 13-14 October, 2000. It will be the 25th congress of our Society and it will mark a substantial period of time during which ESMO has grown steadily as regards number of members, quality of its official journal, *Annals of Oncology*, and relevance of its role in the political issues of oncology in Europe.